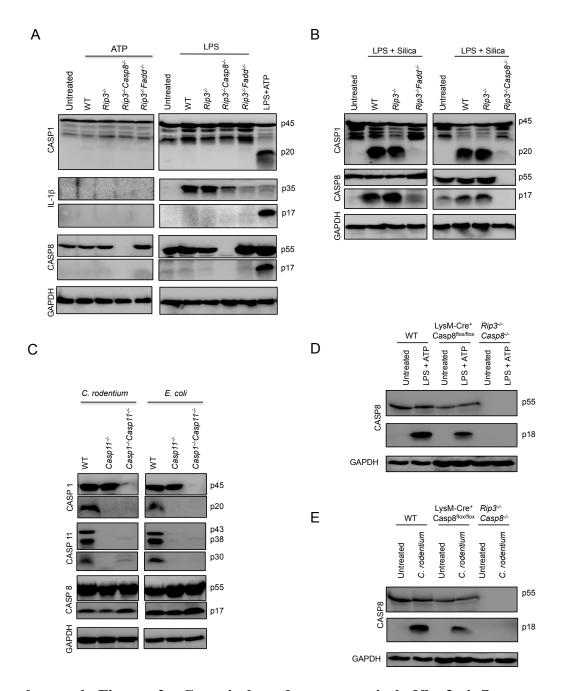
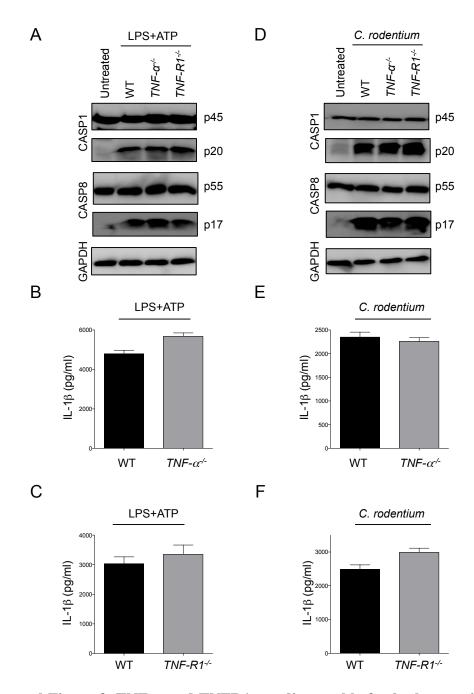


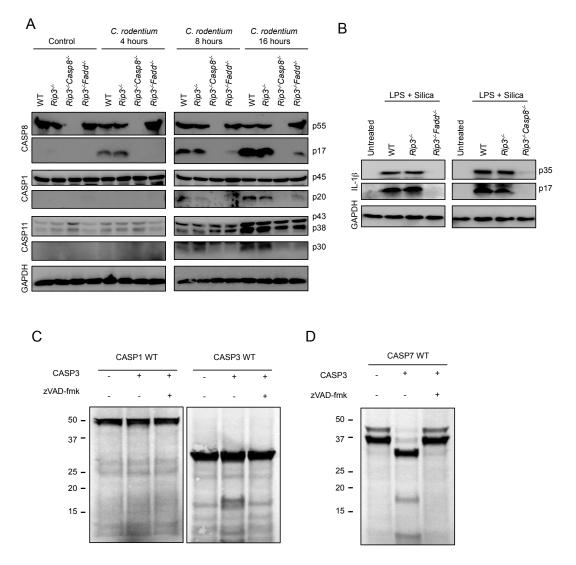
Supplemental Figure 1. Phenotypic and phagyocytic analysis of *Rip3^{-/-}Casp8^{-/-}* and *Rip3^{-/-}Fadd^{-/-}* BMDMs. (A-C) WT, *Rip3^{-/-}Casp8^{-/-}* and *Rip3^{-/-}Fadd^{-/-}* BMDMs were visualized by light microscopy (A) and analyzed for surface expression of the myeloid/macrophage markers CD11b (B) and F4/80 (C) by flow cytometry. (D) WT, *Rip3^{-/-}Casp8^{-/-}* and *Rip3^{-/-}Fadd^{-/-}* BMDMs were either left untreated or stimulated with LPS (20ng/ml) for 24 hours before surface expression of CD86 was analyzed by flow cytometry. (E-G) WT, *Rip3^{-/-}Casp8^{-/-}* and *Rip3^{-/-}Fadd^{-/-}* BMDMs were infected with a GFP-expressing *C. rodentium* strain (Citrobacter-GFP) (E), or incubated with FITC-labeled zymosan (F) or ovalbumin (OVA) (G) for 4 hours before uptake of these particles was determined by flow cytometry. Data are representative of at least 3 independent experiments.



Supplemental Figure 2. Canonical and non-canonical Nlrp3 inflammasome activation. (**A and B**) WT, *Rip3^{-/-}*, *Rip3^{-/-}Casp8^{-/-}* and *Rip3^{-/-}Fadd^{-/-}* BMDMs were either left untreated or stimulated with ATP for 30 minutes (**A, left**), LPS (20ng/ml) for 4 hours (**A, right**) or LPS (20ng/ml) for 3 hours followed by Silica 5 (500µg) for 5 hours (**B**). Cell lystates were then immunoblotted for the indicated proteins. (**C**) WT, *Casp11^{-/-}* and *Casp1^{-/-}Casp11^{-/-}* BMDMs were infected with *C. rodentium* or *E. coli* (m.o.i. 25) for 24 hours as described in Materials and Methods, and lysates were immunoblotted for the indicated proteins. (**D and E**) WT, *LysM-Cre⁺-Casp8^{flox/flox}*, and *Rip3^{-/-}Casp8^{-/-}* BMDMs were left untreated, stimulated with LPS+ATP, or infected with *C. rodentium* as described in Materials and Methods. Cell lysates were immunoblotted for caspase-8 and GAPDH. Data are representative of 3 independent experiments.



Supplemental Figure 3. TNF- α and TNFR1 are dispensable for both canonical and non-canonical Nlrp3 inflammasome activation. (A-F) WT, $TNF-\alpha^{-1}$ and $TNF-R1^{-1}$ BMDMs were stimulated with 20ng/ml LPS for 4 hours (A-C), the last 30 minutes of which in the presence of 5mM ATP or infected with C. rodentium for 24 hours (D-F). Cell lysates were immunoblotted for caspases-1 and -8 (A and D), and culture supernatants were analyzed for IL-1 β (B, C, E and F). Data show mean \pm s.e.m., and are representative of 3 independent experiments.



Supplemental Figure 4. (A) Kinetic analysis of Nlrp3 inflammasome activation by C. rodentium infection. WT, Rip3-/-, Rip3-/-Casp8-/- and Rip3-/-Fadd-/- BMDM cells were stimulated with 25 m.o.i. of C. rodnetium for 4, 8 and 16 hours. Cell lysates were analyzed for indicated proteins by western blot. Data are representative of two independent experiments. (B) LPS+Silica-induced IL-1\beta cleavage in Rip3-\(^{1}\)Fadd-\(^{1}\) **BMDM.** WT, Rip3^{-/-}, Rip3^{-/-}Casp8^{-/-} and Rip3^{-/-}Fadd^{-/-} BMDMs were stimulated with LPS for 3 hours followed by Silica 5 (500µg) for 6 hours. Cell lysates were analyzed for IL-1B induction and cleavage by western blot. (C and D) Cleavage of procaspase-3 and procaspase-7 by recombinant mouse caspase-3 in vitro. (C) Procaspase-1 and procaspase-3 were produced in vitro, and incubated with recombinant caspase-3 (35ng) at 37°C for 1 hour before procaspase-1 and -7 processing was analyzed by autoradiography. In some setups, caspase-3 was pre-incubated with 1µM zVAD-fmk prior to co-incubation with procaspase-1 and -3. (D) Wildtype procaspase-7 was produced in vitro, and incubated with recombinant caspase-3 (35ng) at 37°C for 1 hour before procaspase-7 processing was analyzed by autoradiography. In some setups, caspase-3 was preincubated with 1µM zVAD-fmk prior to co-incubation with procaspase-7.